

The Effect of Some Macrofungi from Turkey Extracts on Cytoplasmic Membrane of Multidrug Resistant Bacteria by Flow Cytometry

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ABSTRACT: In this study; some macrofungi extracts were investigated for their abilities to enhance bacterial permeability by flow cytometry. These experiments exhibited the enhancement of these extracts to disrupt the cytoplasmic membrane of living bacterial (Listeria innocua and Escherichia coli) cells. Antibacterial activity of various macrofungi samples (Pholiota Lucifera, Pleurotus Ostreatus, and Ganoderma Applanatum) against resistant Escherichia Coli and Listeria innocua bacteria was measured by Flow Cytometry. The samples were collected from various regions of Fungi Antakya, Alahan, and Derince of Hatay city of Turkey. These experiments were designed to detect uptake of PI & SYTO by enhancing with a ranged concentration of macrofungi extracts. For this purpose, macrofungi extracts were diluted in the presence of 10% dimethyl sulfoxide in the bacterial culture and incubated at 70 °C for positive control at 70 °C for positive control and at 25 °C for 1 hour for the negative control, and various samples were prepared. According to the results, the macrofungi Ganaderma applanatum, which shows the most antibacterial activity in the negative direction in Listeria innocua bacteria, is the least effective Pleurotus Ostreatus. In Escherichia negative coli bacteria, the most activity while the least is Ganoderma Applanatum, effective is Pholiota Lucifera

KEYWORDS: Macrofungi , flow cytometry, antibacterial effects, Lischeria Innocua, Escherichia Coli.

I. INTRODUCTION

The natural active compounds found in medicinal plants belong to various chemical

structures including polyphenolic compounds, flavonoids, essential oils, and vitamins and some of these compounds have anticancer, antioxidant, and antimicrobial activity. However, these compounds have been little known about mechanisms to confer antibacterial drug resistance. In recent years, research on antimicrobial, anticancer, and antioxidant properties of plant research has increased by researchers. Phenolic compounds, which include functional derivatives such as phenolic acids, flavonoids, anthocyanins, and stilbenes, are functional structures that make up the largest family of all secondary metabolite classes¹. Polyphenols act as antioxidant compounds on oxidative stress. Thus, they play an important role in scavenging reactive oxygen species (ROS), which is another equivalent of toxic free radicals and radicals [1,2]. Many antibacterial studies on various macrofungus species have been reported in the literatüre [3,4]. Pathogenic microorganisms have gained resistance to traditionally used antibacterial drugs. Therefore, the researchers studied plants, fungi, algae etc, and many natural sources investigated their antibacterial activities against microorganisms [15-21]. Naturally, in recent years, antibacterial drug studies and functional food studies related to macrofungi have taken place a lot in the literatüre [22]. It has been reported in the literature that wild mushrooms accumulate a wide variety of pharmacological agents in their bodies. These agents are known to have anticancer, immunostimulating, hypotensive, hypocholesterolemic and antibacterial effects [23-25]. Cultured mycelial macrofungi are prescribed to treat a variety of ailments in many countries, particularly China. The macrofungi species are Phellinus linteus (Teng), Phellinus ignarius (L)

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Quel. and Phellinus robusticus P. Karst. It has an important area of use in China, Phellinus rimosus (Berk.) Pilat. (Hymenochaetaceae), Ganoderma lucidum (Curt: Fr.) P. Karst. (Ganodermataceae) and Navesporus floccosa (Bres.) (Polyporaceae) macrofungi grow on trees. Phellinus rimosus, one of these macrofungi, has antioxidant, antitumor and anti-inflammatory propertie [25-29]. Likewise, this applies to Navesporus floccosa macrofungi. Cortinarius type fungi have about 2000 species in the world and are the most abundant macrofungi in the world. Taxonomic studies have been carried out on such macrofungi in Australia, distinguishing new species [30]. Beattie et al. studied the antibacterial metabolites of fungi of the Cortinarius type [31]. In the Philippines, macrofungi are generally found in the mountainous regions of the country, and approximately 4698 macrofungi species and 1031 of them have been described [32,33]. Studies on the antibacterial activity of wild Basidiomycota and Ascomycota fungi (Cyclocybe aegerita, Cortinarius traganus, Gyroporus castaneus, Neoboletus luridiformis, Rubroboletus Lupinus, Gyromitra esculenta and Helvella crispa) were conducted in France. Of these, the Cyclohexanic extract of Gyromitra esculenta was found to have the strongest efficacy [34[]]. Yamaç et al investigated mycelial and culture antioxidant activities of various mushrooms from Turkey [35]. In these and similar studies, antibacterial and antioxidant activities of many fungi from various countries have been reported [36-41]. In this study, antibacterial activities of some macrofungi (Pholiota Lucifera, Ganoderma Applanatum, and Pleurotus Ostreatus) belonging to some regions in Hatay province were tested against Listeria innocua and Escherichia coli bacteria using flow cytometry test measurements.

II. EXPERIMENTATION

Materials and methods

Macrofungi samples (Pholiota Lucifera, Ganoderma Applanatum, and Pleurotus Ostreatus)were collected by Baba. The voucher specimen is stored in the Fungarium at the Biology Department of Mustafa Kemal University. Pholiota Lucifera was collected from Derince road (200 m). Antakya in Hatay city Derince in Turkey, Ganoderma Applanatum was collected from Alahan road (100 m) Antakya in Hatay city in Turkey, and Pleurotus Ostreatus was collected from

Derince road (100 m) Antakya in Hatay city in Turkey.

Preparation of the methanol extracts

The macrofungi sample, weighing about 100 g was extracted with methanol at 40-45°C for 2 hours (3 times). The filtrates were combined and concentrated in vacuo at 45°C. Finally, the extracts were then lyophilized and kept in the dark at 4 °C until tested.

Extraction can also be done with the supercritical CO_2 method. This method can also be used, especially in order to prevent secondary metabolites from being affected by test conditions [42].

Experimental Stage

Initially optimum conditions were determined. Macrofungi samples (6 mg/ml-3mg/ml-1,5mg/ml, respectively) has been mixed in 10 % dimethyl sulfoxide, respectively. Then 200µL of extract for added to bacterial culture. PI (10%) and SYTO (10%) concentrations were prepared for Listeria innocua and PI (10%) and SYTO concentrations prepared (undiluted) were for Escherichia coli. PBS buffer was used for both Listeria innocua and Escherichia coli. In the experiments for Listeria Innocua bacteria 2.5 µL PI (diluted) and 2.5 µl, SYTO (diluted) were applied in a 500 µl bacteria culture. The samples were incubated in a 1-hour time unit at 70 °C for the positive control. In the negative control, 200 µl of dimethylsulfoxide added. 10% was This application was incubated for a period of 25 ° C for a 1-hour. In experiments for Escherichia coli, 2.5 µL PI (diluted) and 2.5 µl SYTO (undiluted) were applied in a 500 µl bacteria culture and incubated in a 1-hour at 70 ° C for the positive control. In the negative control, 200 µl of 10% dimethylsulfoxide was added and incubated for a 1-hour at 25 °C. Finally, in the presence of 10% dimethylsulfoxide, 6 mg/ml, 3mg/ml, and 1.5mg/ml macrofungi extracts were added separately and incubated at 25 °C for 1 hour, respectively.

III. RESULTS AND DISCUSSION

Taking the positive and negative controls of three species of macrofungi samples (Pholiota Lucifera, Ganoderma Applanatum, and Pleurotus Ostreatus) against Listeria Innocua and Escherichia Coli bacteria, their antibacterial efficacy was demonstrated.



Listeria Innocua tests



Fig. 1 Negative (A) and positive (B) control of Listeria innocua

Pholiota Lucifera (effect of Listeria innocua)



The negative and positive control of macrofungi samples against Listeria Innocua bacteria is shown in fig 1. and the antibacterial activity of Pholiota Lucifera against Listeria Innocua bacteria is shown in fig. 2. According to fig. 2, when the drug concentration is 6 mg/mL, it is understood that 31,90% of the living Listeria innocua cells die and this ratio decreases as the concentration decreases. This ratio is 21,31% while the concentration is 1,5 mg/mL.



concentration is 6 mg/mL, it is understood that

23,94% of the living Listeria innocua cells die and



According to fig. 3, when the drug this ratio of

this ratio decreases as the concentration decreases. This ratio is 23,17% while the concentration is 1,5 mg/mL.



Gnaoderma Applanatum (effect of Listeria innocua)



Fig 4. Percentage of dead cell in Listeria innocua bacterial culture

Gnaoderma Applanatum was a very interesting fungus in this study. In fig. 4, when the drug concentration is 6 mg/mL, it is understood that 99,58% of the living Listeria innocua cells die and

this ratio decreases as the concentration decreases. This ratio is 76,37% while the concentration is 1,5 mg/mL.

	6 mg/mL	3 mg/mL	1,5 mg/mL
Pholiota Lucifera	31,90	23,37	21,31
Pleurotus	23,94	23,74	23,17
Ostreatus			
Gnaoderma	99,58	95,61	76,37
Applanatum			

Escherichia coli tests



Fig. 5 Negative (A) and positive (B) control of Escherichia coli

Pholiota Lucifera (effect of E-coli)



According to fig.6, when the drug concentration is 6 mg/mL, it is understood that 8,67% of the living Listeria innocua cells die and

this ratio decreases as the concentration decreases. This ratio is 5,30% while the concentration is 1,5 mg/mL.



Pleurotus Ostreatus (effect of E-coli)



Fig. 7 Percentage of dead cell in E-coli bacterial culture

Fig. 7 showed that when the drug concentration is 6 mg/mL, it is understood that 9,99% of the living Listeria innocua cells die and this ratio

decreases as the concentration decreases. This ratio is 8,18% while the concentration is 1,5 mg/mL.





Fig. 8 Percentage of dead cell in E-coli bacterial culture

Ganoderma Applanatum showed remarkable results against E-coli bacteria, just like in Listeria innocua. According to fig. 8, when the drug concentration is 6 mg/mL, it is understood that 41,910% of the living Listeria innocua cells die and this ratio decreases as the concentration decreases. This ratio is 16,40% while the concentration is 1,5 mg/mL.

	6 mg/mL	3 mg/mL	1,5 mg/mL
Pholiota Lucifera	8,67	6,18	5,30
Pleurotus Ostreatus	9,99	9,35	8,18
Gnaoderma Applanatum	41,91	24,73	16,40

Listeria monocytogenes is a widespread pathogen that can be found in water, silage, sewage, slaughterhouse waste, cow's milk, human and animal feces. Just like E-coli, it is a bacterium that poses a danger to human health [41]. In this study, especially the effect of Gnaoderma Applanatum on Listeria innocua is quite remarkable. It appears to kill 99.58% of Listeria innocua bacteria at the highest concentration (6mg/mL) (Table 1). It was determined that it was effective at a rate of 41.91% on E-coli. It has been understood that while it has an effect of approximately 20 percent on Pholiota Lucifera and Pleurotus Ostreatus Listeria innocua at all three concentrations, it is not very effective on E-coli and has an effect below ten percent. Looking at these results, Gnaoderma Applanatum showed remarkable differences. With this feature, it is understood that it can have an important function especially in food control and protection and can be evaluated in the food industry.

IV. CONCLUSION

The antibacterial test activity was observed against bacteria in the cytoplasmic membrane (Lischeria Innocua and Escherichia Coli) by flow cytometry of 3 species of macrofungi (Pholiota Lucifera, Pleurotus Ostreatus, and Ganoderma applanatum). In the results obtained, the macrofungi species Ganoderma Applanatum shows the highest negative efficiency in Lischeria

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Innocua and Escherichia Coli bacteria. The macrofungi species that show the least negative activity in Lischeria Innocua bacteria is the Pleurotus Ostreatus genus. In Escherichia Coli bacteria, the macrofungi genus showing the least negative activity is Pholitota Lucifer. In antibacterial activity tests in these macrofungi species, it showed a more resistant property against Escherichia Coli bacteria against Lischeria Innocua bacteria.

REFERENCES

- García-Pérez P, Lozano-Milo E, Landín M, Gallego PP. Combining Medicinal Plant In Vitro Culture with Machine Learning Technologies for Maximizing the Production of Phenolic Compounds. Antioxidants. 9 (3) (2020) 210.
- [2]. Garcia-Perez P, Lozano-Milo E, Losada-Barreiro S, Bravo-Diaz C. Plant Antioxidations in Food Emulsions. In: In Some New Aspects of Colloidal Systems in Food. Milani: IntechOpen; 2018:11–29.
- [3]. Silva DD De, Rapior S, Sudarman E, et al. Bioactive metabolites from macrofungi: Ethnopharmacology, biological activities and chemistry. Fungal Divers, 62(1) (2013) 1–40.
- [4]. Haider K, Trojanowski J. A Comparison of the Degradation of 14C-Labeled DHP and Corn stalk lignins by Micro- and Macrofungi and Bacteria. In: Lignin Biodegradation: Microbiology, Chemistry, and Potential Applications. CRC Press; (2019) 111–134.
- [5]. Karaca B, Çöleri Cihan A, Akata I, Altuner EM. Anti-Biofilm and Antimicrobial Activities of Five Edible and Medicinal Macrofungi Samples on Some Biofilm Producing Multi Drug Resistant Enterococcus Strains. Turkish J. Agric. - Food Sci. Technol., 8(1) (2020) 69.
- [6]. Chandra Deka A. Antimicrobial Properties and Phytochemical Screening of Some Wild Macrofungi of Rani-Garbhanga Reserve Forest Area of Assam, India (2017).
- [7]. Lu H, Lou H, Hu J, Liu Z, Chen Q. Macrofungi: A review of cultivation strategies, bioactivity, and application of mushrooms. Compr. Rev. Food Sci. Food Saf., 19(5) (2020) 2333–2356.
- [8]. Godwin ME. Macrofungal Extracts on the Bacteria Inhibition of Bacillus subtilis Amruta Ponugupati. Eur. J. Heal. Biol. Educ. 4(1) (2015) 1–8.
- [9]. Liu Y, Sun Q, Li J, Lian B. Bacterial diversity among the fruit bodies of ectomycorrhizal and saprophytic fungi and their corresponding hyphosphere soils. Sci. Rep. 8(1) (2018) 1–10.
- [10]. Sheena N, Ajith TA, Mathew AT, Janardhanan

KK. Antibacterial activity of three macrofungi, Ganoderma lucidum, Navesporus floccosa and Phellinus rimosus occurring in South India. Pharm. Biol. 41(8) (2003) 564–567.

- [11]. Bala N, Aitken EAB, Fechner N, Cusack A, Steadman KJ. Evaluation of antibacterial activity of Australian basidiomycetous macrofungi using a high-throughput 96-well plate assay. Pharm. Biol. 49(5) (2011) 492– 500.
- [12]. Yu G, Sun Y, Han H, et al. Coculture, An Efficient Biotechnology for Mining the Biosynthesis Potential of Macrofungi via Interspecies Interactions. Front. Microbiol. 12 (2021) 663924.
- [13]. E FO, Oyetayo O V, Imisioluwa Awala S, Awala SI. Evaluation of the mycochemical composition and antimicrobial potency of wild macrofungus, Rigidoporus microporus (Sw) Mushroom Enrichment View project Assessment of nutraceutical properties of bioactives in underutilized macrofungi in Nigeria. View project Evaluation of the mycochemical composition and antimicrobial potency of wild macrofungus, Rigidoporus microporus (Sw). (2017).
- [14]. Ali S. Anti-pathogenic efficacy of Indian edible macrofungi Dacryopinax spathularia (Schwein) and Schizophyllum commune (Fries) against some human pathogenic bacteriae. (2019)
- [15]. Karalija E, Parić A, Dahija S, Bešta-Gajević R, Ćavar Zeljković S. Phenolic compounds and bioactive properties of Verbascum glabratum subsp. bosnense (K. Malý) Murb., an endemic plant species. Nat. Prod. Res. 34(16) (2020) 2407–2411.
- [16]. Najmeh Aboutorabi S, Nasiriboroumand M, Mohammadi P, Sheibani H, Barani H. Preparation of antibacterial cotton wound dressing by green synthesis silver nanoparticles using mullein leaves extract. J. Renew. Mater. 7(8) (2019) 787–794.
- [17]. Al-Snafi AE. Iraqi Medicinal Plants with Antibacterial Effect-A Review Clinically tested medicinal plants View project Medicinal plants with cardiovascular effects View project Iraqi Medicinal Plants with Antibacterial Effect-A Review (2019)
- [18]. Singh Sandhu S, Kumar S, Prasad Aharwal R. Isolation and Identification of endophytic fungi from Ricinus communis Linn. and their antibacterial activity Studies on Bioactive Compounds from Endophytic Fungi as Prospective antimicrobial Agents View project Evaluation of economic traits of Sesame View project. (2016).

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- [19]. Xu L, Meng W, Cao C, et al. Antibacterial and antifungal compounds from marine fungi. Mar. Drugs. 13(6) (2015) 3479–3513.
- [20]. Sandsdalen E, Haug T, Stensvåg K, Styrvold OB. The antibacterial effect of a polyhydroxylated fucophlorethol from the marine brown alga, Fucus vesiculosus. World J. Microbiol. Biotechnol. 19(8) (2003) 777– 782.
- [21]. Holten Lützhøft HC, Halling-Sørensen B, Jørgensen SE. Algal toxicity of antibacterial agents applied in Danish fish farming. Arch. Environ. Contam. Toxicol. 36(1) (1999) 1–6.
- [22]. Poucheret P, Fons F, Rapior S. Biological and Pharmacological Activity of Higher Fungi: 20-Year Retrospective Analysis. (1998).
- [23]. Silva DD De, Rapior S, Sudarman E, et al. Bioactive metabolites from macrofungi: ethnopharmacology, biological activities and chemistry 62 (2013) 1-40.
- [24]. Sun T, Tang J, Powers JR. Antioxidant activity and quality of asparagus affected by microwave-circulated water combination and conventional sterilization. Food Chem. 100(2) (2007) 813–819.
- [25]. Nowacka N, Nowak R, Drozd M, et al. Antibacterial, antiradical potential and phenolic compounds of thirty-one polish mushrooms. PLoS One. 10(10) (2015) e0140355.
- [26]. Ikekawa T, Nakanishi M, Uehara N, Chihara G, Fukuoka F. Antitumor action of some Basidiomycetes, especially Phllinus inteus. Gann, Japanese J. Cancer Res. (1968;59(2):155–157.
- [27]. Khatun S. Research on Mushroom as a Potential Source of Nutraceuticals: A Review on Indian Perspective. Am. J. Exp. Agric. 2(1) (2012) 47–73.
- [28]. Ajith T., Janardhanan K. Antioxidant and antiinflammatory activities of methanol extract of Phellinus rimosus (Berk) Pilat (2001).
- [29]. Ajith TA, Jose N, Janardhanan KK. Amelioration of cisplatin induced nephrotoxicity in mice by ethyl acetate extract of a polypore fungus, Phellinus rimosus. J. Exp. Clin. Cancer Res. 21(2) (2002) 213–217.
- [30]. Garnica S, Weiß M, Oertel B, Oberwinkler F. A framework for a phylogenetic classification in the genus Cortinarius (Basidiomycota, Agaricales) derived from morphological and molecular data. Can. J. Bot. 83(11) (2005) 1457–1477.
- [31]. Beattie KD, Rouf R, Gander L, et al. Antibacterial metabolites from Australian macrofungi from the genus Cortinarius.

Phytochemistry. 71(8–9) (2010) 948–955.

- [32]. Parlucha JA, Soriano JKR, Yabes MD, Pampolina NM, Tadiosa ER. Species and functional diversity of macrofungi from protected areas in mountain forest ecosystems of Southern Luzon, Philippines. Trop. Ecol. (2021) 1:3.
- [33]. Cruz TEE Dela, Kuhn R V, Javier AOM, Parra CM, Quimio TH. Status of the Myxomycete Collection at the UPLB-Museum of Natural History (UPLB-MNH) Mycological Herbarium. Philipp. J. Syst. Biol. (2009) 3(1).
- [34]. Morel S, Vitou M, Masnou A, et al. Antibacterial activity of wild mushrooms from france. Int. J. Med. Mushrooms. 23(1) (2021) 79–89.
- [35]. Börühan Çetin M, Ceylan G, Cantürk Z, et al. Screening of Antioxidant Activity of Mycelia and Culture Liquids of Fungi from Turkey. Microbiol. Russian Fed. 90(1) (2021) 133– 143.
- [36]. Badalyan SM, Borhani A. Medicinal, nutritional, and cosmetic values of macrofungi distributed in mazandaran province of northern iran (Review). Int. J. Med. Mushrooms. 21(11) (2019) 1099–1106..
- [37]. Nacua AE, Pacis HJM, Manalo JR, et al. Short communication: Macrofungal diversity in MT. Makiling forest reserve, Laguna, Philippines: With floristic update on roadside samples in Makiling botanic gardens (MBG). Biodiversitas. 19(4) (2018) 1579–1585.
- [38]. Kinge TR, Goldman G, Jacobs A, Ndiritu GG, Gryzenhout M. A first checklist of macrofungi for South Africa. MycoKeys. 63 (2020) 1–48.
- [39]. Hassan F, Ni S, Becker TL, et al. Evaluation of the antibacterial activity of 75 mushrooms collected in the vicinity of oxford, ohio (Usa). Int. J. Med. Mushrooms. 21(2) (2019)131– 141.
- [40]. Doğan HH, Duman R, Özkalp B, Aydin S. Antimicrobial activities of some mushrooms in Turkey. Pharm. Biol. 51(6) (2013) 707–711.
- [41]. Yavuz M, Korukluoğlu M. Listeria monocytogenes'in Gıdalardaki Önemi ve İnsan Sağlığı Üzerine Etkileri The Importance of Listeria monocytogenes in Foods and Its Effect on Human Health. 24(1) (2010) 1–10.
- [42]. Nuralin L., Tosun A., Erol F., Akgun F. B., Gürü M, Optimisation of quercetin amount via response surface methodology for elaeagnus seeds by supercritical CO2 extraction method. J. of The Faculty of Eng. And Arc.of Gazi Unv. (2017), 32(4), 1233-1241.